

BIOSORPTION OF SOME HEAVY METALS FROM CRUDE OIL POLLUTED SOIL USING BANANA PEELS AND GINGER WASTE AS BIOSORBENT.

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ABSTRACT

In this study biosorption of some selected heavy metals from crude oil polluted soil was carried out using banana peels and ginger waste as biosorbent. The heavy metals analyzed are zinc, chromium, cadmium, lead and nickel. The polluted soil sample was collected from Ubeji community and the absorbent were gotten from fruit vendors and Igbudu market. The crude oil polluted soil collected from Ubeji community was digested using wet acid digestion method and analysed using the atomic absorption spectrophotometer (AAS) to determine the concentration of the various heavy metals and the result gotten were as follows 16.55 mg/kg, <0.10 mg/kg, <0.50 mg/kg, 17.00mg/kg, <0.10 for zinc, chromium, cadmium, lead and nickel respectively, the concentration of the heavy metals were later analysed after treatment by the biosorbent (banana peel and ginger waste), the concentration gotten for the first week after treatment are 13.05mg/kg, <0.10mg/kg, 0.50mg/kg, 14.00mg/kg, <0.10mg/kg for zinc, chromium, cadmium, lead and nickel respectively for banana peel (50g) and 12.97mg/kg, <0.10mg/kg, 0.50mg/kg, 14.90mg/kg, <0.10mg/kg for zinc, chromium, cadmium, lead and nickel respectively for ginger waste (50g), Also for 100g of adsorbent the concentration gotten for the first week after treatment are 10.05mg/kg, <0.10mg/kg, 0.50mg/kg, 12.30mg/kg, <0.10mg/kg for zinc, chromium, cadmium, lead and nickel respectively for banana peel (100g) and 10.65mg/kg, <0.10mg/kg, 0.50mg/kg, 15.05mg/kg, <0.10mg/kg for zinc, chromium, cadmium, lead and nickel respectively for ginger waste (100g). Significant differences were noted for the subsequent weeks. Comparing the result of concentration of the heavy metals before treatment and after treatment for 50g and 100g of the absorbent, it clearly shows that banana peels and ginger waste were effective in reducing the concentration of zinc, chromium, cadmium, lead and nickel in the crude oil polluted soil sample and also 100g of biomass was found to be more efficient than 50g of the biomass, as such the biosorbent can be used for the bioremediation of heavy metals in crude oil polluted soil.

INTRODUCTION

Crude oil is a naturally-occurring resource which contains hydrocarbon deposits and other organic materials (Tamuno and Felix, 2006). There is an estimated consumption of 90 million barrels of oil daily worldwide. The use of crude oil and its fractions has a negative impact on the environment, damaging ecosystems through spills and releasing a range of pollutants in the environment (Guerrio, Pokhilko, Fernandez, Haliday, Millar, & Hillston, 2012).

In Nigeria, oil spill has become a common occurrence. An estimated 9 million to 13 million barrels have been spilled since oil drilling started in Nigeria in 1958 (Kadafa, 2012). The Niger Delta region which covers about 70,000km² of wetlands is a home to more than 20 million people and 40 ethnic groups (Agochi, 2014; Ugbomeh and Atubi, 2010). There is the slow poisoning and destruction of vegetation and agricultural land by oil spills. Idoniboye (2000) reported that oil spills presented far more hazardous effects on soil and soil organisms than air because whatever plants take in goes into the food chain and water cycle. Over the years, different methods have been applied in cleaning up crude oil polluted soils and of all these methods, bio-related clean up types appeared as the most desirable approach due to its low cost and ability to hinder the accumulation of contaminants (Esin and Ayten, 2011).

Bio-sorption is a property of certain types of inactive dead microbial biomass to bind and concentrate contaminants from even very dilute aqueous solutions. It has advantage over the other conventional

treatment methods, as it is low cost, highly efficient, minimal chemical and biological sludge; no need of additional nutrients and has the possibility to regenerate sorbents and possible metal recovery and most times waste materials are used.

Crude oil pollution seems inevitable in areas where crude exploration/refining are carried out. This spill not only cause ecological damage, but destroys the livelihood of inhabitants of the area and also their health. Besides the devastating effects it has on the environment, it also incurs huge cost of cleaning up the environment. As the current techniques for cleanup are capital intensive. This necessitated the search for cheaper means of clean ups. Not only that these clean up techniques are expensive, they are also environmentally friendly. These agricultural by product are widely available and are of little or no economic value and some of them in fact present a disposal problem. Moreover, dead cells are cheaper, effective in reducing heavy metals to very low levels and requires less care and maintenance

Also, the dead biomass could be easily regenerated and reused (Wankasi et al 2005, Horsfall et al 2006). The reason for the use bio sorption over conventional treatment methods is that it is cost effective, has high efficiency, minimization of chemicals and other biological sludge and the possibility of metals recovery.

The aim of the study is to examine the bio-sorption of heavy metals from oil polluted soil, Nigeria using Banana and Ginger peels *in order* to evaluate the concentration of these heavy metals in soil from the study area and to determine the concentration of these heavy metals in soil after treatment (applying remediation).

This study covers the use of agricultural waste (dead biomass like banana and ginger wastes) as adsorbent for the treatment of some heavy metal (zinc, chromium, cadmium, lead, nickel,) contaminant in crude oil polluted soil collected from Ubeji community, Warri south local government area of Delta State.

MATERIAL AND METHODS

The Banana peels were collected from the fruit vendors and the garlic waste was gotten from food vendors and also purchased from the market. The Banana peels and the ginger were sorted and dried for 10 days. After the moisture contents have been removed, the complex was dried in the oven at 120 degree for 2 hours and was grinded. then sieved through a 2mm mesh size of a micro sieve.

The crude oil polluted soil sample was collected at Ubeji community Delta State. Ubeji community is situated at geographical position of coordinates 5.572326 and 5.702848. The soil was collected with a spade to about 15cm in depth and the sample was poured into a polythene bag. The soil samples were collected randomly at different parts of the community and mixed together. Control soil was collected about 500m away from spill site. The soil samples were air dried for 24 hours, and sieved to pass through 2mm mesh and stored at room temperature prior to chemical analysis. The experimental pots were set up using 50g and 100g of the biomass to 200g of soil and were labeled accordingly.

After 7 days, 20 g of the crude oil polluted soil was prepared for digestion and was taken to the Atomic absorption spectrophotometer (AAS) machine for analyses of the various heavy metals (cadmium, zinc, lead, chromium nickel)

RESULT AND DISCUSSION

Table1 showing the result of adsorption rate of the adsorbents on zinc

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (50g)	16.55	13.05	10.37	9.55	8.00	7.25
GINGER WASTE (50g)	16.55	12.97	11.01	9.76	8.89	8.10

Zinc Adsorption

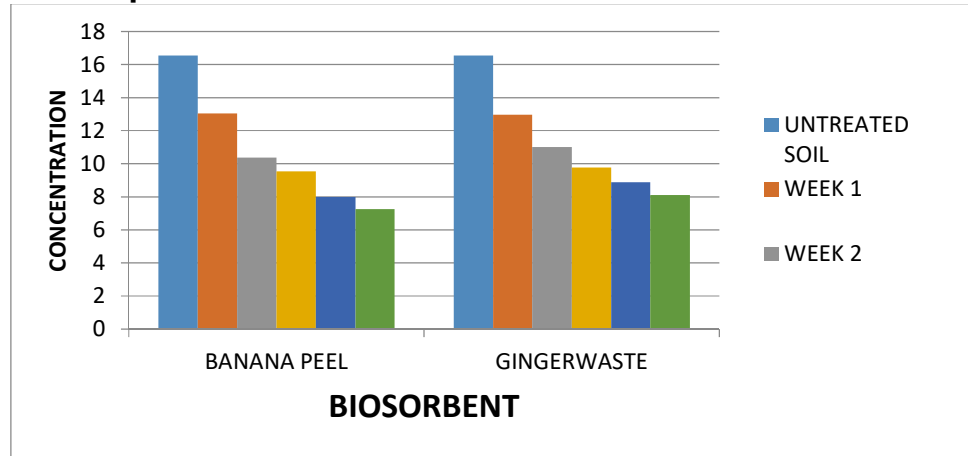


Figure 1 Zinc Adsorption

Table 1 above illustrate the reduction of zinc concentration from 16.55 to 13.05 for week one, 10.37 for week two, 9.55 for week three, 8.00 for week four and 7.25 for week five by banana peel (50g) while ginger waste (50g) reduced the concentration from 16.55 to 12.97 for week one, 11.01 for week two, 9.76 for week three, 8.89 for week four and 8.10 for week 5.

Table 2 showing the result of adsorption rate of the adsorbents on chromium

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (50g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (50g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10

Table 2 shows the result of chromium concentration which was constant at <0.10 (indicating that there is no significant level of chromium concentration in the soil sample) for the five weeks for both banana peel (50g) and ginger waste (50g).

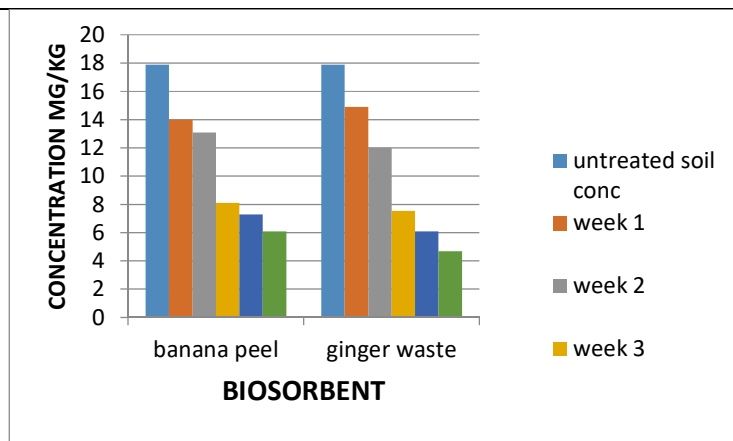
Table 3 showing the result of adsorption rate of the adsorbents on cadmium

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (50g)	<0.50	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (50g)	<0.50	<0.10	<0.10	<0.10	<0.10	<0.10

Table 3 shows the result of cadmium concentration which was constant at <0.10 (indicating that there is no significant level of cadmium concentration in the soil sample) for the five weeks for both banana peel (50g) and ginger waste (50g).

Table 4 showing the result of adsorption rate of the adsorbents on lead

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (50g)	17.90	14.00	13.10	8.10	7.30	6.09
GINGER WASTE (50g)	17.90	14.90	12.00	7.55	6.10	4.70



Lead Adsorption
Figure 2 Lead Adsorption

Table 4 illustrate the reduction of lead concentration from 17.90 to 14.00 for week one, 13.10 for week two, 8.10 for week three, 7.30 for week four and 6.09 for week five by banana peel (5g) while ginger waste (50g) reduced the concentration from 17.90 to 14.90 for week one, 12.00 for week two, 7.55 for week three, 6.10 for week four and 4.70 for week 5.

Table 5 showing the result of adsorption rate of the adsorbents on nickel

Adsorbent	Untreated soil sample conc	Week 1	Week 2	Week 3	Week 4	Week 5
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		(mg/kg)	Treated soil sample conc (mg/kg)	Treated soil sample conc (mg/kg)	Treated soil sample conc (mg/kg)	Treated soil sample conc (mg/kg)	Treated soil sample conc (mg/kg)
BANANA (50g)	PEEL	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER (50g)	WASTE	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10

Table 5 shows the result of nickel concentration which was constant at <0.10 (indicating that there is no significant level of cadmium concentration in the soil sample) for the five weeks for both banana peel (50g) and ginger waste (50g).

Table 6 showing the result of adsorption rate of the adsorbents on zinc

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc mg/kg	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (100g)	16.55	10.05	9.02	5.0	1.10	<0.10
GINGER WASTE (100g)	16.55	10.65	7.50	2.75	<0.10	<0.10

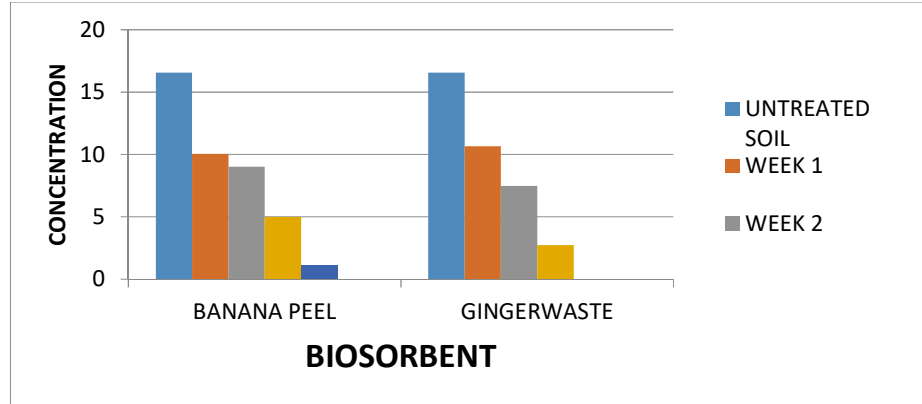


Figure 3 Zinc Adsorption

Table 6 illustrate the reduction of zinc concentration from 16.55 to 10.05 for week one, 9.02 for week two, 5.00 for week three, 1.10 for week four and <0.10 for week five by banana peel (100g) while ginger waste (100g) reduced the concentration from 16.55 to 10.65 for week one, 7.50 for week two, 2.75 for week three, <0.10 for week four and <0.10 for week 5.

Table 7 showing the result of adsorption rate of the adsorbents on chromium

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (100g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (100g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10

Table 7 shows the result of chromium concentration which was constant at <0.10 (indicating that there is no significant level of chromium concentration in the soil sample) for the five weeks for both banana peel (100g) and ginger waste (100g).

Table 8 showing the result of adsorption rate of the adsorbents on cadmium

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (100g)	<0.50	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (100g)	<0.50	<0.10	<0.10	<0.10	<0.10	<0.10

Table 8 shows the result of cadmium concentration which was constant at <0.10 (indicating that there is no significant level of cadmium concentration in the soil sample) for the five weeks for both banana peel (100g) and ginger waste (100g).

Table 9 showing the result of adsorption rate of the adsorbents on lead

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc (mg/kg)	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (100g)	17.90	12.30	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (100g)	17.90	15.05	<0.10	<0.10	<0.10	<0.10

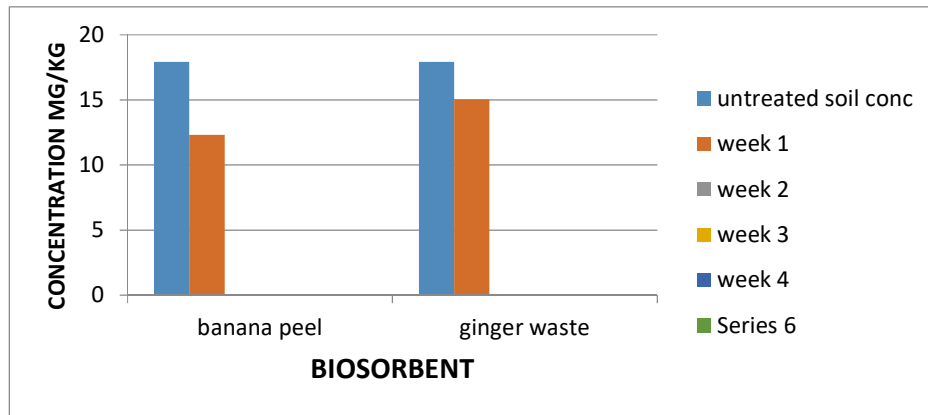


Figure 4 Lead Adsorption

Table 9 illustrate the reduction of lead concentration from 17.90 to 12.30 for week one, <0.10 for week two, <0.10 for week three, <0.10 for week four and <0.10 for week five by banana peel (100g) while ginger waste (100g) reduced the concentration from 17.90 to 15.05 for week one, <0.10 for week two, <0.10 for week three, <0.10 for week four and <0.10 for week 5.

Table 10 showing the result of adsorption rate of the adsorbents on nickel

Adsorbent	Untreated soil sample conc (mg/kg)	Week 1 Treated soil sample conc (mg/kg)	Week 2 Treated soil sample conc (mg/kg)	Week 3 Treated soil sample conc mg/kg	Week 4 Treated soil sample conc (mg/kg)	Week 5 Treated soil sample conc (mg/kg)
BANANA PEEL (100g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
GINGER WASTE (100g)	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10

Table 10 shows the result of nickel concentration which was constant at <0.10 (indicating that there is no significant level of cadmium concentration in the soil sample) for the five weeks for both banana peel (100g) and ginger waste (100g).

Comparative Discussion of Result for 50g and 100g of Biomass

ZINC

Comparing the result gotten for 50g of biomass and 100g of biomass, it show that adsorption of heavy metals is more efficient in 100g of banana peel and ginger waste than 50g Of banana peels and ginger waste as the concentration of zinc was reduced from 16.55 to 7.25 for banana peel(50g) and the concentration of zinc was reduced from 16.55 to 8.10 for ginger waste (50g), while the concentration of zinc was reduced from 16.55 to <0.10 using banana peel (100g) for the five week and the concentration of zinc was reduced from 16.55 to <0.10 using ginger waste(100g) in five week.

LEAD

Comparing the result of gotten for 50g of biomass and 100g of biomass, it show that adsorption of heavy metals is more efficient in 100g of banana peel and ginger waste than 50g Of banana peels and ginger waste as the concentration of lead was reduced from 17.90 to 6.09 for banana peel (50g) and the concentration of lead was reduced from 17.90 to 4.70 for ginger waste (50g), while the concentration of lead was reduced from 17.90 to <0.10 using banana peel (100g) in two week

and the concentration of zinc was reduced from 17.90 to <0.10 using ginger waste (100g) in two week.

CHROMIUM

The concentration of chromium was constant at <0.10 mg/kg(indicating that there is no significant level of chromium) for both the banana peel (50g) and ginger waste (50g) for the five week. Also for 100g of the biomass the concentration of chromium was constant at <0.10 mg/kg (indicating that there is no significant level of chromium) for both the banana peel (100g) and ginger waste (100g) for the five week.

CADMIUM

The concentration of cadmium was constant at <0.10 mg/kg (indicating that there is no significant level of chromium) for both the banana peel (50g) and ginger waste (50g) for the five week. Also for 100g of the biomass the concentration of cadmium was constant at <0.10 mg/kg (indicating that there is no significant level of chromium) for both the banana peel (100g) and ginger waste (100g) for the five week

NICKEL

The concentration of nickel was constant at <0.10 mg/kg (indicating that there is no significant level of chromium) for both the banana peel (50g) and ginger waste (50g) for the five week. Also for 100g of the biomass the concentration of nickel was constant at <0.10 mg/kg (indicating that there is no significant level of chromium) for both the banana peel (100g) and ginger waste (100g) for the five week

CONCLUSION

The treatment of Crude oil polluted soil using agricultural waste such as Ginger waste and Banana Peels has been studied and found feasible for both 50g and 100g of the biomass Adsorption of each heavy metal varies with the nature of the adsorbents and the 100g of biomass was seen to more efficient in adsorbing the heavy metals.

Results from this study can be used to access the suitability of this agricultural waste as adsorbents for heavy metal removal (bioremediation) or reduction of concentration as a cheap alternative to the commercially available expensive activated carbon.

RECOMMENDATION

Efficiency can certainly be improved and more research is recommended in this area, Research on other agricultural waste using the same method can help in analyzing the effectiveness of the process. Adsorption capacity on concentration of heavy metals was examined with the different agricultural waste for five weeks. Although, there are various other parameters whose effect could be investigated and results analyzed to make an even better assessment if time permit.

REFERENCES

- Abel-Salam A.F; Shalenda M; Elaby and Jehian B. Ali (2014).Antimicrobial and antioxidant activities of red onions, garlic and leeth in soilsage.African journal of microbiology research vol. 8 (27).pp.2574-2582. 2 July, 2014. Doi:10.5897/AJMR 2014.6755..
- Agochi, O.C, (2014). Gas flaring in the Niger Delta Nigeria: An act of inhumanity to man & his environment. World Academy of Science, Engineering and Technology.International Journal of Social, Behaviour, Educational, Economic, Business & Industrial Engineering.Vol: 8, No; 7, 2014.

- Akpan, E.E., Kingsley .C. and Nwadinigwe.C.A. (2013).Bioremediation of Hydrocarbon polluted soil in the lowland forest ecosystem in the Niger Delta through enhanced natural Attenuation process (ENAPP). International Journal of Applied Science and Technology.ISSN 2221-0997 (print), 2221-1004 (online).
- Ayangbenro, S.A. and BabalolaO.O . (2007). A new strategy for heavy metal polluted environments: A review of microbial Bisorbents. Int. J. Environ. Res.public health 2017, 14, 94: 10:3390/ijerph 401 40094. www.mdpl.com/journal/ijerph.
- Ayotamuno, M.J; Kogbara, R.B; Ogaj, S.O.T. and Probert S.D (2006). Bioremediation of a crude oil polluted agricultural soil at Port Harcourt Nigeria. Applied energy, 83:1249-1257.
- Baird J. "Oil's shame in Africa". Newsweek, 156 (4).27-27, 2010.
- Eludoyin, O.S and Oghenetega O.J (2015). Temporal remediation of crude oil contaminated soil using remediation by enhanced attenuation (RENA) technique in Mbodo, Community, Port Harcourt, Nigeria. Tropical Agriculture (Trinidad) 93 (i), 36-46, 2016.
- EsinEradogan and AytenKaraca (2011). Bioremediation of crude oil polluted soil. Asian journal of biotechnology 3(3): 206-213, 2011.
- Faisal, M. and Hasnain, S. (2004). 'Microbial conversion of Cr (VI) into Cr (III) in industrial effluent', African Journal of Biotechnology, vol. 3, No. 11,,pp 610-617.
- Gadd, G.M. (2002). Bioremedial potential of microbial mechanisms of metal mobilization and immobilization.CurropinBiotechnol, 200 June; 11(3): 27-9.
- Gadd, G.M. (2004). Microbial influence on metal mobility and application for Bioremediation.Geodema 122(2-4) :109-119 Oct 2004. Doi:10.1016/J. Geoderma. 2004.01.002.
- Gadd, G.M. (2007) Geiomyecology: biogeochemical transformations of rocks, minerals, metals &radionuclides by fungi, bioweathering and bioremediation. Mycol Res 111, 3-49.
- Geoffery M. Gadd and Marina Fomina (2011).Uranium and Fungi.Geomicrobiology Journal vol. 28, 155 5-6,2011.
- Geoffrey Michael Gadd (2008); Biosorption! Critical Review of scientific rationale environmental importance and significance for pollution treatment.Published online in Wielyinterscience 29 July 2008. (www.interscience.com D01 10. 1002/jctb.1999
- Grath S. P., Chaudri A. M., and Giller K. E.(1994). Summary 15th World Congress of Soil Science, Acapulco, México.
- Gune, A; Alpasalan, M and Inal, A (2014). Plant growth and fertilizer.Ankara University of Agriculture publication. No: 1539, Ankara, Turkey.
- Guerrio, M.L., Pokhilko, A., Fernandez, A.P., Haliday, K.J., Millar, A.J., Hillston, J. (2012).Stochastic properties of plant circadian clock. J.R. SOC. Interface 9 744-756. Published 31 August 2011.DOI:10.1098/rsif.2011.0378.

- Hammaini, A; Gonzalez, F; Ballester, A; Blazquez, M.L. and Munoz, J.A. (2003). 'Simultaneous uptake of metals by activated sludge', Mineral Engineering, vol. 16, No. 8, pp 723-729.
- Hamzah, A., Phan, C. W., Abu Bakar, N. F., and Wong, K. K. (2013). Biodegradation of crude oil by constructed bacterial consortia and the constituent single bacteria isolated from Malaysia. Bioremediat. J.17, 1-10.
- Hanaor, D.A.H., Ghadiri, M.; Chrzanowski, W; Gan, Y; (2014) ."Scalable Surface Area characterization by ElectroKinetic Analysis of Complex Anions Adsorption" (PDF). Langmuir. 30 (50): 15143-15152. Doi: 10. 1021/1a503581e PMD 25495551
- James B Condon, (2012). Surface Area and Porosity Determination by Physiosorption.Measuerement and Theory.Elesvier.
- Jiang C. Y., Sheng X. F., Qian M., and Wang Q. Y. (2008). Isolation and characterization of heavy metal resistant Burkholderia species from heavy metal contaminated paddy field soil and its potential in promoting plant growth and heavy metal accumulation in metal polluted soil. Chemosphere; 72:157–164.
- JomovaK, and Valka M (2011): Advances in metal induced oxidative stress and human disease. Toxicol; 283 (2-3): 65-87.
- Joshi N. 2003. Biosorption of heavy metals.AnM.Sc in Biotechnology.Thesis. Department of Biotechnology & Environmental sciences, Thapar Institute of Engineering & Technology, Patiala 14 7004, may 2003.
- Kadafa A.A. (2012). "Environmental Impacts of Oil Exploration and Exploitation in the Niger Delta of Nigeria". Global Journal of Science frontier research environment and earth science vol. 12, issue 3. Version10.2012.
- Kakulu S.E. and Osibanjo O, 1992. Pollution studies of Nigerian Rivers; Trace metal levels of surface water in the Niger Delta area Int. J. Environn. 124 (1-3) : 263-275.
- Kanmani P., Aravind J., and Preston D. (2012).Remediation of chromium contaminants using bacteria.International Journal of Environmental Science and Technology; 9:183– 193.
- Katircioglu, H: Aslim, B, Yuksekdao Z.N; Mercan. N; and Beyatli (2003).Production of poly – hydroxybutyrate (PHB) and differentiation of putative Bacillus mutant strains by SDS-PAGE of total cell protein. Afr. J. Biotechnol; 2: 147-149.
- Lannoy, G. De (2001).Vegetable crops in Tropical Africa 395-511. In: Raemaeks R.H. (Editor). Crop production in Tropical Africa.1540 pp. Directorate General for International Co-operation {DGIC} Belgium.
- Lee Y. C., and Chang S. P. (2011).The bio-sorption of heavy metals from aqueous solution by Spirogyra and Cladophora filamentous macroalgae. Bioresource Technology; 102 (9) 5297–5304.
- Li, H; Shi, W; Shao, H. and Shao M. (2009).The remediation of Leadpolluted garden soil by natural zeolite; Journal of Harzardous materials, vol. 169, Nos. 1-3, pp 1106-1111.

- Lin H.V; Rogulja, A; Cadigan, K.M. (2004). Wingless eliminates ommatidia from the edge of the developing eye through activation of apoptosis .development 131 (10): 2409-2418.
- Machado M. D., Soares E. V., and Soares H. M. (2010).Removal of heavy metals using a brewer's yeast strain of *Saccharomyces cerevisiae*: chemical speciation as a tool in the prediction and improving of treatment efficiency of real electroplating effluents. *Journal of Haz-ardous Materials*; 180(1–3) 347–353.
- Macleon J.L; Dawe D.C; Hardy B, and Hettel G.P. (eds). (2002). *Rice almanac* (3rd edition).Philippines, IRRI, WARDA, CIAT and FAO.
- NIV CH, Volesky B, Ueiman D (2007). Bisorption of arsenic (V) with a cid-washed crab shells. *Water res. Jun*; 41(11) : 24 73-8.
- Obi .I. Chinedu (2012).Oil spill disaster in Nigeria – The Borga field oil spill disaster.Published in AWE International magazine, June 2012, Edition. www.aweimagazine.com.
- Olayiwola O.A, Agbolade J.O. and Oladapo A.J (2013).Daily exposure of road side top soil to Lead at Olaiya Roundabout, Oshogbo, Nigeria. *Int. J. Eng. Sci. Investment*. 2(5):55-60.
- Olayiwola, O.M. (2011). Development and validation of a model for longitudinal survey optimum response.PhD thesis, University of Ibadan, Nigeria.
- OlobaniyiS.B., and Efe S.I. (2006) comparative analysis of rain water and ground water quantityin an oil producing area of Nigeria: *J. Enviro. Health Res*; 6(2): 111-117.
- OlomukoroJ.O., and Azubuike C.N. (2009). Heavy metals and mar invertebrate communities in bottom sediments of Ekpan Creek, Warri, Nigeria: *Jordan J. Biol.sci.*, 2(1): 1-8.
- Onifade, A.K., Abubakar, F.A. and Ekunday, F.O. (2007).Bioremediation of crude oil polluted soil in the Niger Delta area of Nigeria using enhanced natural Attenuation.*Research Journal of Applied Sciences* 2007 volume 2, Issue 4, Pp 498-504.
- Onyekwelu, J.C., Mosandl R. &Stimm, B. (2008): Tree species diversity and soil status of primary and degraded tropical rainforest ecosystems in South-western Nigeria. *Journal of Tropical Forest Science* 20(3): 193–204.
- Orji, F.A., Abiye, A. I and Dike, E.N. (2012).Laboratory scale bioremediation of petroleum hydrocarbon - polluted mangrove swamps in the Niger Delta using cow dung. *Malaysian Journal of Microbiology*, Vol 8(4), pp. 219-228.
- Osuji, L.C (2002). Some environmental hazards of oil pollution in the Niger Delta, Nigeria. *Africa. Interdisc. Stud*. 3(i) 11-12.
- Osuyi, L.C. and Onojake, C.M. (2004). Trace heavy metals in crude oil; A case study of Ebocha-8 oil spill polluted site in Niger Delta Nigeria. *Chem. Biodiv*. 1: 1708-1715.
- Oyibo (2013).Phytoremediationof some tropical soils contaminated within petroleum crude oil. A PhD thesis- University of Ghana 2013.<http://dtidleharder.net/12345678915177>.

- Pandey, P.K; Choubey, S; Verma, Y; Pandey M; Kamal, S.S.K. and Chandrashekhar, K. (2007).Biosorptive removal of Ni (II) from waste and industrial effluent, International Journal of environmental research and public health, vol. 4, No. 4, pp 332-339.
- Pelletier, E. Delille, D. and Delille, B. (2004). "Crude oil bioremediation in sub-Antarctic intertidal sediments: chemistry and toxicity of oiled residues," Marine Environmental Research, vol. 57, no. 4, pp. 311–327.
- Rajeswari K, Subashkumar R, Vijay Graman K. degradation of Textile Dyes by Isolated *Lysinibacillus sphaericus* strain RSV-1 and *Stenotrophomonas maltophilia* strain RSV-2 and toxicity Assessment of degraded products. J. Environ Anal Toxicol 2014;4:4.
- Roane T. M., Josephson K. L., and Pepper I. L. (2001). Dual-bioaugmentation strategy to enhance remediation of contaminated soil. Applied and Environmental Microbiology; 67 (7) 3208–3215.
- Salman H. Abbas, Ibrahim. M. Ismail, Tarek. M. Mostafa and Abbas H. Salaymon, (2014).Biosorption of heavy metals .A review Journal of chemical science & Technology.Vol.3 ISS. 4 pp 74-102.
- Sari F; Braus G.H; Irrniger S (2007). A process Independent of the anaphase-promoting complex contributes to instability of the yeast S phase cyclin C1b5. J Biol-Chem 2007 Sep. 7; 282 (36) 26614-22. Epub 2007 July 9.
- Say R., Yimaz N., and Denizli A. (2003). Removal of heavy metal ions using the fungus *Penicillium canescens*. Adsorption Science and Technology; 21 (7) 643–650.
- Shao, H.B; Chu, L.Y; Ruan,, C.J; Li, H; Guo, D.G. and Li, W.X. (2010). 'Understanding molecular mechanism for improving phytoremediation of heavy metal contaminated soil', critical review of Biotechnology, vol. 30, No. 1, pp 23-30.
- Sizmur, T; Palumbo-Roe, B; Watts, M.J; Hodson, M.E. (2011). Impact of the earthworm *Lumbricus terrestris*
- Tamuno S. and Felix J.M. (2006). Crude Oil: A blessing or a curse- The case of Niger Delta .Journal of research in National development. Vol. 4(2) 2006):pp 53-58.
- Tastan B. E., Ertugrul S., and Donmez G. (2010).Effective bio-removal of reactive dye and heavy metals by *Aspergillus versicolor*.Bioresource Technology; 101(3) 870–876.
- Tokar, E.J; Benbrahim-Tallaa, L. and Waalkes, M.P (2011).Metal ions in human cancer development.Metal ions on life science, 8; 375-401.
- Ugbomeh B.A; and Atubi A.O (2010).Preliminary multivariate analysis of the factors of socio-economic development of Nigeria- A case study of Delta State Nigeria.African research review- vol. 4, No. 4 2010.<http://dx.doi.org/10.431A/affreav. v.414-69222>.
- United States Army Corps of Engineers (2003).Agriculturally Based Bioremediation of Petroleum-Contaminated Soils and Shallow Groundwater in Pacific Island Ecosystems.
- Uzoije, A.P and Sgunwamba, J.C (2011).Physiochemical properties of soil in Relation to varying rates of crude oil pollution. Journal of environmental science & technology, 4:313-323.

- Veiria, R.H. and Volesky (2000).Biosorption: a solution to pollution? IntMicrobiol. 2000 mar; 3 (I): 17-24
- Venosa A.D. and Zhu X. (2003). "Biodegradation of crude oil contaminating marine shorelines and freshwater wetlands," Spill Science and Technology Bulletin, vol. 8, no. 2, pp. 163– 178.
- Wang J and Chen C. (2006). Bio-sorption of heavy metals by saccharonylescerevisiae: a review. Biotechnol Adv. 2006 Sep-oct; 24(5) 427-51.Epub 2006 Jun 5.
- Wang J. and Chen .C. (2009).Bio-sorbents for Heavy metal removal & their future. Biotechnology advances 27(2): 195-226. March 2009 Doi: 10:1016 IJ. Biotechadv. 2008.11.002. source Pub med.
- Wankasi D.; Horsfall M. Jnr, and Spiff A.I. (2005). Desorption of Pb^{2+} and Cu^{2+} from Nipa palm (Nypafriuticansklurmb) biomass. African Journal of Biotechnology vol. 4(9), pp 923-927. ISSN 1684-5315.
- Wankasi D.; Horsfall M. Jnr, and Spiff A.I. (2006). Sorption kinetics of Pb^{2+} and Cu^{2+} ions from aqueous solution by Nipa palm (Nypafriuticansklurmb) shoot biomass. Electronic Journal of Biotechnology vol. 9, No. 5, pp 587-592.
- Williams, J.J; Seaton, N.A; Duren, T. (2011). Influence of surface groups on the diffusion of gases in MCM-41: A molecular Dynamics study. Journal of Physical Chemistry.Vol. 115.Doi: 10. 1021/JP112073z.
- Wuara, R.A and Okiemute F.E (2011).Heavy metal in contaminated soils. A review of sources, chemistry, Risks and best available strategies for remediate. Communications in soil science & plant analysis, 42, 111-222.<http://dx.doi.org/10.5402/2011/40264>
- Xin Q. G; Pan W. B; Zhang T. P. (2003). On phytoremediation of heavy metal contaminated soils. Ecologic science 22 (3): 275-299.